

original report

Toward Electronic Surveillance of Invasive Mold Diseases in Hematology-Oncology Patients: An Expert System Combining Natural Language Processing of Chest Computed Tomography Reports, Microbiology, and Antifungal Drug Data

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abstract **Purpose** Prospective epidemiologic surveillance of invasive mold disease (IMD) in hematology patients is hampered by the absence of a reliable laboratory prompt. This study develops an expert system for electronic surveillance of IMD that combines probabilities using natural language processing (NLP) of computed tomography (CT) reports with microbiology and antifungal drug data to improve prediction of IMD.

Methods Microbiology indicators and antifungal drug-dispensing data were extracted from hospital information systems at three tertiary hospitals for 123 hematology-oncology patients. Of this group, 64 case patients had 26 probable/proven IMD according to international definitions, and 59 patients were uninfected controls. Derived probabilities from NLP combined with medical expertise identified patients at high likelihood of IMD, with remaining patients processed by a machine-learning classifier trained on all available features.

Results Compared with the baseline text classifier, the expert system that incorporated the best performing algorithm (naïve Bayes) improved specificity from 50.8% (95% CI, 37.5% to 64.1%) to 74.6% (95% CI, 61.6% to 85.0%), reducing false positives by 48% from 29 to 15; improved sensitivity slightly from 96.9% (95% CI, 89.2% to 99.6%) to 98.4% (95% CI, 91.6% to 100%); and improved receiver operating characteristic area from 73.9% (95% CI, 67.1% to 80.6%) to 92.8% (95% CI, 88% to 97.5%).

Conclusion An expert system that uses multiple sources of data (CT reports, microbiology, antifungal drug dispensing) is a promising approach to continuous prospective surveillance of IMD in the hospital, and demonstrates reduced false notifications (positives) compared with NLP of CT reports alone. Our expert system could provide decision support for IMD surveillance, which is critical to antifungal stewardship and improving supportive care in cancer.

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INTRODUCTION

Invasive mold disease (IMD) has significant health and economic consequences¹⁻³ and is likely to increase as the population of immunocompromised patients expands.⁴ IMD most often presents as pneumonia in patients with impaired immunity due to a variety of causes, including chemotherapy for cancer, immunomodulatory drugs, or transplantation.^{3,5,6} IMD is associated with mortality rates of > 50% in patients with hematologic malignancies and transplantation recipients^{3,6}

and may adversely affect long-term cancer cure rates due to modifications in curative chemotherapy regimens.⁷ Hospitals spend millions on antifungal drugs but have not invested in prospectively monitoring their local epidemiology (ie, institution-specific pathogens, rates, and patients affected), despite it being a key performance indicator of antifungal drug stewardship programs.⁸ Knowledge of local fungal epidemiology is important for several reasons. It informs local guidelines,^{8,9} infection prevention efforts, clinical trial design,

clinical audit, and benchmarking that addresses interfacility clinical variation.¹⁰ Epidemiologic surveillance of IMD is difficult because of the absence of a consistent laboratory prompt, such as a positive blood culture. As a result, case finding relies on manual interrogation of multiple data sources, including bedside clinical review, radiology, and microbiology, which makes it a costly, onerous task not performed by hospitals outside research protocols.^{3,5,6} As a result of an absence of surveillance data, hospitals are ill equipped to evaluate their clinical practice, rationalize antifungal drug use, detect outbreaks, or identify new, previously under-recognized at-risk patient groups.

To facilitate monitoring of IMD in hospitals, we developed natural language processing (NLP) of chest computed tomography (CT) reports.^{11,12} We have previously shown that the primary screening method for IMD, with a high sensitivity to maximize case finding, should focus on chest CT.^{11,12} CT is universally performed when IMD is suspected, and pulmonary involvement is present in 90% to 100% of patients with IMD.^{3,5,13} Although lung sampling (ie, biopsy or bronchoalveolar lavage [BAL]) and other laboratory indicators (eg, sputum culture, biomarkers) could be used for epidemiologic surveillance, they are not performed with the same frequency as CT scans.

Our baseline text classifier, detailed elsewhere,^{11,12} was based on automatically learned, hierarchically organized, cascading text categorization techniques. In brief, we obtained 1,880 free-text CT reports for 270 patients with IMD and 257 control patients from three tertiary hospitals.¹¹ We analyzed IMD evidence at patient, report, and sentence levels. Training data were obtained by three infectious disease physicians who annotated a subgroup of 449 reports from 79 patients with IMD and 68 control patients for language features suggestive of IMD.¹¹ We tested a variety of machine learning, rule-based, and hybrid systems, with feature types including bags of words, bags of phrases, and bags of concepts, as well as report-level structured features.^{11,12} The best system (using support vector machines) achieved high recall over unseen data, with a sensitivity of 91%, specificity of 79%, and receiver operating characteristic (ROC) area of 0.92, with few clinically significant missed notifications (0.9%) at report level,¹¹ whereas in a separate experiment that used a slightly smaller data set ($n = 1,716$ free-text CT reports), it identified 100% of patients with IMD.¹² With a negative predictive value of 97% to 99% (across a range of hypothetical IMD prevalence rates), IMD could be excluded with confidence.¹¹ Our next goal was to minimize

false notifications (ie, improve specificity) while maintaining or improving case detection (ie, sensitivity). Therefore, we developed an expert system that incorporated derived probabilities from NLP of CT reports, medical knowledge rules, and machine learning classifiers for processing adjunctive data (microbiology and antifungal drug-dispensing data) to enable real-time, sustainable, and network-wide surveillance of IMD with minimal effort.

METHODS

Study Design and Setting

This was a retrospective case-control cohort study of patients from three tertiary adult university-affiliated hospitals (Alfred Health, Peter MacCallum Cancer Institute, and Royal Melbourne Hospital). Alfred Health and Royal Melbourne Hospital operate statewide allogeneic and autologous hematopoietic stem-cell transplantation (HSCT) services, which collectively perform approximately 250 allogeneic transplantations per year. Peter MacCallum Cancer Institute performs autologous HSCT only. The human research ethics committees at each site approved the study.

Clinical Data and Definitions

We selected a random convenience sample of 64 infected case patients and 59 uninfected control patients from the original data set used to develop the text classifier.¹¹ Microbiology and antifungal drug-dispensing data for each clinical encounter (defined from admission to discharge, death, or transfer from hospital) were extracted from hospital information systems and combined with probabilistic outputs of the text classifier derived from the earlier study.¹¹

IMD was classified according to internationally accepted definitions.¹⁴ Possible infections had suggestive radiologic features in the appropriate clinical context but lacked positive microbiology, whereas probable or proven infections had positive microbiology, such as isolation of a fungal pathogen from sputum or tissue.¹⁴

Microbiology included microscopy and culture of specimens from sterile and nonsterile body sites, including blood, BAL, tissue, pleural fluid, sputum, and biomarkers (galactomannan [GM], panfungal or *Aspergillus*-specific polymerase chain reaction [PCR]). We obtained pharmacy-dispensing data for the following antifungal drugs—fluconazole, voriconazole, posaconazole, liposomal amphotericin, caspofungin, itraconazole, and terbinafine—as well as the dosage, formulation (oral, intravenous), presentation (vials, tablets), and dates dispensed. None of the hospitals had electronic prescribing

systems in place, so drug-dispensing data approximated the actual dose administered to the patient.

Development of the Expert System

Our approach can be deconstructed into two phases: 1) We detected the patients at high likelihood of IMD by combining the probabilistic output of the text classifier with expert knowledge rules about IMD, and 2) for the remaining patients, we trained a classifier on all the available features including output of the text classifier, antifungal drug-dispensing data, and microbiology investigations and results.

Preprocessing

Preprocessing involved transforming the manually collected data from long to wide format. Each electronic event (ie, CT scan, drug dispensed, microbiology result) per clinical encounter was given a time stamp (date). Electronic events represented the following variables: probabilistic outputs of text classification of CT scan reports derived from the earlier study,¹¹ results of key microbiological investigations, and antifungal drug use (Table 1). Categorical variables (ie, positive, negative, or no test available) were assigned for each electronic event.

For example, electronic events were named Scan_Any positive, Blood_Any positive, BAL_Any

positive, Pleural Fluid_Any positive, PCR_Any positive, GM_Any positive, Sterile_Any positive, and Non_sterile_Any positive, with “Any positive” denoting all positive results in that category for each clinical encounter. Scan_Any positive was set to positive if the text classifier returned a value ≥ 0.5 with the highest probability from any scan in the clinical encounter supplied to the expert system. Isolation of *Candida* species from the pulmonary or oropharyngeal tract was not regarded as significant.

A clinical encounter with a negative CT scan by NLP and positive sterile site microbiology (ie, Sterile_Any positive) was marked as positive for IMD and not sent to the final classifier. Isolation of fungal pathogens from a nonsterile site, such as sputum, usually denotes colonization rather than invasive infection. However, for purposes of epidemiologic surveillance, we regarded the isolation of molds from sputum (ie, Non_Sterile_Any positive) as significant because of the high likelihood of either occult or subsequent invasive disease in this high-risk population.¹⁵

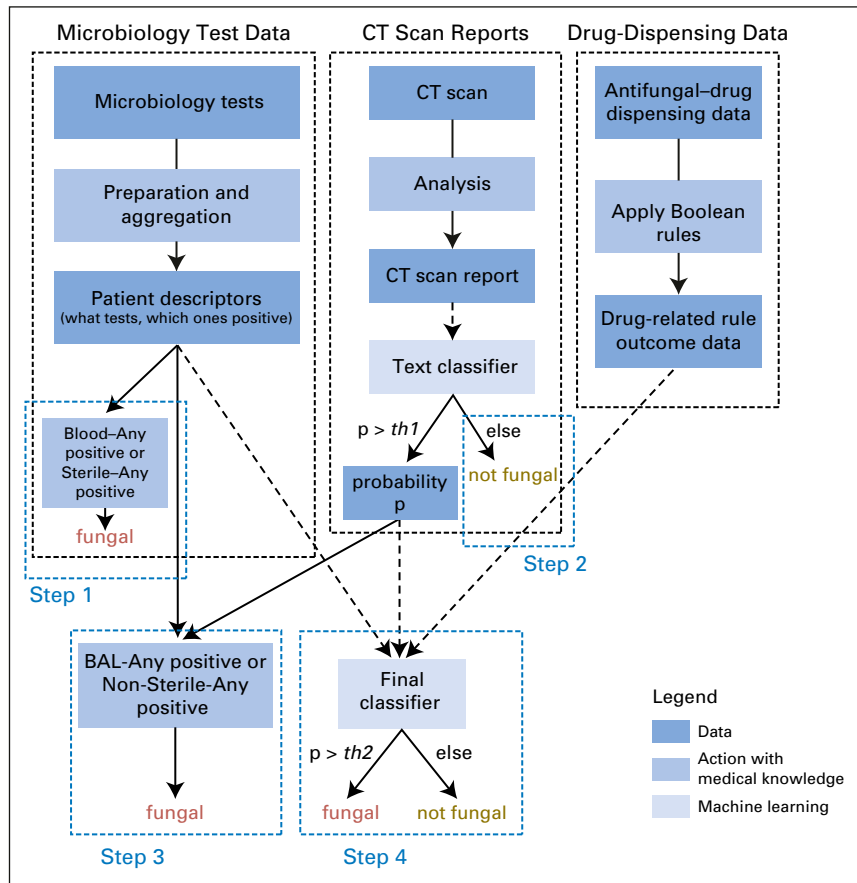
We created handcrafted rules on the basis of medical expertise (M.R.A.-R., K.T., M.S.) and informed by real-world data from a previous study¹ to interpret complex antifungal drug-dispensing data. These rules aimed to identify drug regimens more likely to reflect definitive treatment of IMD

Table 1. Definitions of Electronic Event Variables for CT, Microbiology, and Antifungal Drug-Dispensing Data Used by the Expert System

| Electronic Event | Hospital Information System | Description |
|-------------------------|-----------------------------|---|
| Scan_Any positive | Radiology | CT scan report with a probability > 0.5 for IMD by the baseline text classifier any time during the patient's clinical encounter (ie, episode of care). |
| Specimen_Any positive | Microbiology | Specimens can be from sterile or nonsterile anatomic sites. These may be samples from blood, tissue, pleural fluid, sputum, or wound sites. Any positive refers to isolation of a fungal pathogen at any time during the patient's clinical encounter (ie, episode of care). |
| Sterile_Any positive | Microbiology | Sterile sites include blood, pleural fluid, BAL, tissue (eg, lung or sinus biopsy). Any positive refers to isolation of a fungal pathogen at any time during the patient's clinical encounter (ie, episode of care). |
| Nonsterile_Any positive | Microbiology | Nonsterile sites include sputum, nasopharyngeal swab, mouthswill, or occasionally, wounds. Any positive refers to isolation of a fungal pathogen at any time during the patient's clinical encounter (ie, episode of care). |
| PCR or GM_Any positive | Microbiology | PCR and GM are molecular biomarkers for fungal pathogens. PCR may be performed on a variety of specimens, including blood and tissue. GM is usually performed on lung specimens (eg, BAL) or serum. Any positive refers to a positive result at any time during the patient's clinical encounter (ie, episode of care). |
| Rule 1 | Pharmacy | Set to “yes” if voriconazole, irrespective of formulation, exceeded 4,200 mg per clinical encounter. |
| Rule 2 | Pharmacy | Set to “yes” if at least 150 mg of liposomal amphotericin was dispensed daily for 7 days or more during a clinical encounter. |
| Rule 3 | Pharmacy | Set to “yes” if 50 mg or more of caspofungin was dispensed daily for 5 days or more per clinical encounter |

Abbreviations: BAL, bronchoalveolar lavage; CT, computed tomography; GM, galactomannan; IMD, invasive mold disease; PCR, polymerase chain reaction.

Fig 1. Architecture of the expert system incorporating natural language processing, microbiology, and antifungal drug-dispensing data for invasive mold disease classification. Sterile specimens include blood, pleural fluid, bronchoalveolar lavage (BAL), and tissue (eg, lung or sinus biopsy). Nonsterile specimens include sputum, nasopharyngeal swab, mouthswab, or occasionally, wounds. Any positive refers to isolation of a fungal pathogen at any time during the patient's clinical encounter (ie, episode of care). Th1 refers to the threshold of 0.5 that was prespecified for the baseline text classifier,^{11,12} meaning that a report with a probability ≥ 0.5 was flagged positive. Th2 is the threshold selected by the best-performing final classifier (Table 3) to maintain sensitivity and maximize specificity. Dashed lines represent inputs to the classifiers as opposed to processing steps and inputs to the expert system. CT, computed tomography.



rather than empirical or prophylaxis indications. Our earlier study¹ observed that median duration of inpatient therapy was 6 days for voriconazole, 7.5 days for liposomal amphotericin, and 6 days for caspofungin in patients with invasive fungal disease. Our drug rules were based on these observations but modified (Table 1), because drug dispensing from pharmacy to the hospital wards is not always consecutive, and the volumes of drug dispensed are variable. If drugs were given but not above the thresholds set by the rules, the variables were set to either “low” or “no.” We did not include rules for fluconazole, itraconazole, and posaconazole, because these drugs are more frequently used for prophylaxis rather than treatment of established IMD, and our pharmacy dispensing data do not provide clinical indications for prescription. Terbinafine was not included in our rules because it is reserved for the rare episode of invasive scedosporiosis, where it is coadministered with voriconazole, a scenario that would have been detected by our voriconazole rule.

Architecture of the Expert System

The expert system (Fig 1) comprised machine-learning classifiers for text analysis and final

classification, algorithms informed by medical knowledge, and clinical data. Fungal isolates from sterile sites, including blood (ie, Blood_Any positive or Sterile_Any positive), which unequivocally represented invasive disease were identified with a medical rule (Step 1, Fig 1). The baseline text classifier then screened all CT reports (Step 2, Fig 1). Positive IMD cases from the text classifier were classified as positive, without sending them to the final classifier, if either BAL_Any positive (ie, fungal isolation from lung) or Non_Sterile_Any positive (ie, fungal isolation from sputum) were labeled as positive. In this way, only undecided cases with negative or no tests on Blood_Any positive, BAL_Any positive, Sterile_Any positive, and Non_Sterile_Any positive were sent to the final classifier (Fig 1). Thus the final classifier processed probabilistic outputs from the text classifier of CT reports, drug data not subject to the aforementioned rules, and negative or missing microbiology.

Classification Algorithms

We experimented with several machine-learning algorithms, shown in Table 2. For implementation, we used the R programming language, together with Weka and the R Weka¹⁶ package. All algorithms

Table 2. Parameters Used for the Machine Learning Algorithms

| Algorithm | Parameters |
|---------------|--|
| Naïve Bayes | — |
| Logistic | MaxIts = -1, ridge = 1.0×10^{-8} |
| Random forest | numTrees = 10, numFeatures = 5 |
| J48 | confidenceFactor = 0.25, minNumObj = 2 |
| SMO | c = 1, cachesize = 250,007, exponent = 1.0 |

Abbreviation: SMO, sequential minimal optimization.

used their Weka default configurations (the C4.5 implementation in Weka is called J48).

Evaluation Procedure

Evaluation was performed for the expert system overall. We used 10-fold cross-validation that used all available data to train and test the expert system, meaning that we obtained a prediction for each of the 123 data points. The results of the expert system for each clinical encounter (with IMD diagnosed or not) was compared with physician-adjudicated opinion on the basis of published diagnostic criteria,¹⁴ which allowed for the calculation of sensitivity, specificity, and ROC.

RESULTS

Patient Characteristics

The data set of 123 patients comprised 64 (52%) patients with IMD and 59 (48%) uninfected control patients (Table 4). Neutropenia ($\leq 0.5 \times 10^9$ cells/L) was present in 86% and 71% of case and control patients, respectively, and was prolonged (median, 17 and 18 days, respectively). A history of HSCT was present in 53% and 46% of case and control patients, and was allogeneic in 85% and 78% of patients, respectively. IMD was probable or proven in 26 of 64 patients (41%). Sinus and/or pulmonary disease occurred in 90% of case patients. Invasive aspergillosis comprised 14 of 26

(54%) of microbiologically confirmed cases, with rare molds, including *Scedosporium* and *Rhizopus* species, identified in nine of 26 (35%).

Performance of the Final Classifier

Performance of the final classifiers for different threshold values is shown in Table 3 and Figure 2. Performance was best using naïve Bayes, which increased the specificity from 50.8% (95% CI, 37.5% to 64.1%) to 74.6% (95% CI, 61.6% to 85.0%), with an ROC of 92.8% (95% CI, 88.0% to 97.5%). With this approach, false notifications fell 48%, from 29 to 15, with improvements in sensitivity from 96.9% (95% CI, 89.2% to 99.6%) to 98.4% (95% CI, 91.6% to 100%).

Error Analysis

Error analysis of the expert system, as shown in Table 5, focused on the naïve Bayes algorithm, because this had the best performance characteristics in experiments. We found one missed patient with IMD (false negative) resulting in an overall missed notification rate of 0.8% (one of 123). This patient had a previous lung resection for *Scedosporium apiospermum* infection in an earlier clinical encounter, but the CT scan for the current encounter was appropriately labeled negative by the text classifier because it represented postoperative changes. Although the patient received liposomal amphotericin and voriconazole sequentially, neither exceeded the thresholds to trigger the drug rules for that particular encounter.

There were 15 patients flagged positive by the expert system (false positives) but subsequently not diagnosed with IMD (Table 5). In all cases, the probability of fungal disease according to the text classifier (Scan_Any positive) was well above the prespecified threshold of 0.5. In other words, the baseline text classifier was responsible for the

Table 3. Performance Characteristics of the Classifiers Compared With the Baseline Text Classifier

| Algorithm | Threshold* | Sensitivity, % (95% CI) | Specificity, % (95% CI) | ROC, % (95% CI) |
|--------------------------|------------|-------------------------|-------------------------|---------------------|
| Baseline text classifier | 0.00 | 96.9 (89.2 to 99.6) | 50.8 (37.5 to 64.1) | 73.9 (67.1 to 80.6) |
| Naïve Bayes | 0.32 | 98.4 (91.6 to 100.0) | 74.6 (61.6 to 85.0) | 92.8 (88.0 to 97.5) |
| Random forest | 0.12 | 96.9 (89.2 to 99.6) | 72.9 (59.7 to 83.6) | 94.1 (89.8 to 98.3) |
| Logistic | 0.00 | 98.4 (91.6 to 100.0) | 50.8 (37.5 to 64.1) | 91.1 (85.7 to 96.5) |
| SMO | 0.12 | 96.9 (89.2 to 99.6) | 69.5 (56.1 to 80.8) | 92.4 (87.5 to 97.3) |
| J48 | 0.10 | 96.9 (89.2 to 99.6) | 57.6 (44.1 to 70.4) | 87.2 (80.6 to 93.7) |

Abbreviations: ROC, receiver operating characteristic; SMO, sequential minimal optimization.

*The algorithm generated a threshold automatically to maintain sensitivity at least as high as the baseline text classifier while maximizing specificity.

Table 4. Characteristics of Patients With and Without IMD

| Characteristic | IMD Group | Control Group |
|--|---------------|---------------|
| No. of patients | 64 (52) | 59 (48) |
| Male sex | 39 (61) | 27 (46) |
| Mean age, years (range) | 53 (24-89) | 51 (18-89) |
| Underlying disease | | |
| AML | 24 (38) | 30 (51) |
| ALL | 10 (16) | 12 (20) |
| Lymphoma | 14 (22) | 7 (12) |
| Chronic leukemia | 6 (9.4) | 1 (1.7) |
| MDS/transformed MDS | 6 (9.4) | 2 (3.4) |
| Multiple myeloma | 2 (3.1) | 3 (5.1) |
| Other | 2 (3.1) | 4 (6.8) |
| Neutropenia (\leq 0.5 cells/L) present | 55 (86) | 42 (71) |
| Median duration of neutropenia, days (IQR) | 17 (13-26) | 18 (10-25) |
| HSCT | 34 (53) | 27 (46) |
| Allogeneic | 29 of 34 (85) | 21 of 27 (78) |
| Autologous | 5 of 34 (15) | 6 of 27 (22) |
| Characteristic of IMDs (n = 64) | | NA |
| Probable and/or proven IMDs | 26 (41) | |
| Possible IMDs | 38 (59) | |
| Site of infection | | |
| Lung | 54 (84) | |
| Sinopulmonary | 3 (4.8) | |
| Sinus | 1 (1.6) | |
| Hepatosplenic | 2 (3.2) | |
| Disseminated | 4 (6.3) | |
| Organism | | |
| <i>Aspergillus fumigatus</i> | 9 | |
| Nonfumigatus <i>Aspergillus</i> species (<i>A. niger</i> , <i>A. flavus</i>) | 2 | |
| Fungal hyphae resembling <i>Aspergillus</i> species | 3 | |
| <i>Scedosporium</i> species | 4 | |
| Any positive PCR | 4 | |
| <i>Rhizopus</i> species | 3 | |
| Other molds (<i>Acrophialophora</i> <i>fusispora</i> , <i>Paecilomyces lilacinus</i>) | 2 | |
| <i>Candida glabrata</i> (coinfection with <i>Scedosporium prolificans</i> fungemia) | 1 | |

NOTE. Data presented as No. (%), except as otherwise indicated.

Abbreviations: ALL, acute lymphoblastic leukemia; AML, acute myeloid leukemia; HSCT, hematopoietic stem-cell transplantation; IMD, invasive mold disease; IQR, interquartile range; MDS, myelodysplastic syndrome; NA, not applicable; PCR, polymerase chain reaction.

majority of false positives; a drug rule was positive in nine encounters, and one encounter had a positive microbiology indicator being Non_Sterile_Any positive (ie, *Aspergillus fumigatus* isolation from

sputum) in a patient with multiple myeloma who did not have an IMD.

DISCUSSION

Our expert system is a promising model for electronic surveillance of IMD in hospitals that extends earlier efforts to detect IMD from chest CT reports using NLP.^{11,12} High sensitivity maximized case finding, and we reduced false notifications (ie, improved specificity) by combining microbiology and antifungal drug-dispensing data with NLP of CT scan reports.^{11,12} Compared with the baseline text classifier, the best performing expert system improved sensitivity slightly from 96.9% to 98.4%. The greatest benefit was seen in specificity, which increased from 50.8% to 74.6%, reducing false notifications by 48% from 29 to 15. Overall accuracy was good, as reflected by area under the ROC, which increased from 73.9% to 92.8% compared with the baseline text classifier alone.

The use of NLP was motivated by the poor sensitivity of other screening approaches for epidemiologic surveillance of IMD. Laboratory-based surveillance of IMD that uses culture and histology is insensitive because microbiology for *Aspergillus* and hyaline molds is positive in \leq 50% of cases,¹⁷ and patients are often too unwell to undergo invasive diagnostic procedures. Molecular biomarkers, including GM or PCR, are not widely available and have a suboptimal sensitivity,^{18,19} which is further reduced by concomitant antifungal therapy administered at the earliest clinical suspicion of IMD.²⁰ Coding data are unreliable for IMD surveillance²¹ and neither timely nor informative enough for outbreak detection. Our expert system that incorporates NLP of CT reports could overcome many of these barriers. It leverages routinely available electronic clinical and operational data that will become easier to access in the computational environment of the electronic medical record (EMR); of note, this work is not dependent on the EMR, which was not present in the study hospitals.

Recent approaches to electronic surveillance of a range of diseases from hospitals have shown promise. A major study demonstrated superiority of NLP of the EMR for surveillance of postoperative complications compared with the more widely used diagnostic codes (eg, sensitivity 82% v 38% for acute renal failure).²² NLP was recognized to have the added benefit of timeliness and the potential to detect while the patient is still in the hospital,^{22,23} a characteristic also relevant to IMD because of the risk of hospital acquisition and/or outbreaks associated with these infections.²⁴

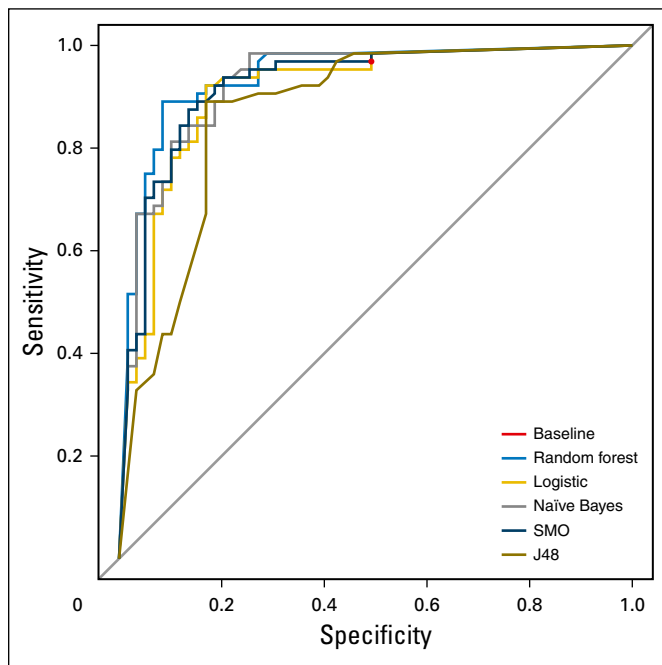


Fig 2. Receiver operating curve of the final classifiers processing probabilistic output from natural language processing of computed tomography reports by the baseline text classifier, microbiology, and antifungal drug dispensing. The red dot represents performance of the baseline text classifier, which had a fixed prespecified threshold of 0.5^{11,12} and a relatively high sensitivity without the addition of microbiology and antifungal drug-dispensing data. SMO, sequential minimal optimization.

The quality of source data is another challenge of electronic surveillance. Antifungal drug-dispensing data were least valuable, partially because of their format (pharmacy dispensing is not equivalent to bedside electronic prescribing), and they lacked the specific reasons for drug choice (ie, prophylaxis, empirical, pre-emptive, or targeted therapy). An audit of a computerized decision support system (CDSS) for antifungal drug prescriptions in pediatric hematology-oncology patients demonstrated the superiority of CDSS for inpatient fungal surveillance over coding data,²⁵ but its usefulness is entirely dependent on prescriber adherence (ie, where prescribers enter their justification for drug choice) with the CDSS.

Other approaches to disease surveillance have combined diagnostic codes with NLP.^{26,27} For hepatocellular carcinoma, diagnostic codes were used as a first level of identification, with NLP subsequently applied to pathology and radiology reports to improve case ascertainment.²⁷ For pneumonia, DeLisle et al²⁸ combined a text classifier with EMR-derived structured data that included clinical features and diagnostic codes. The addition of text classification of chest radiograph reports increased the positive predictive value of EMR-based case-detection algorithms by 20% to 70%, while retaining sensitivities of 58% to 75%.²⁸ Ahmed et al²⁹ used a combination of search term queries of an EMR along with extraction of vital signs, medications, and laboratory values to identify a variety of acute conditions including sepsis, pneumonia, aspiration, acute

pancreatitis, and shock, which are risk factors for acute respiratory distress syndrome. For sepsis, they observed that an automated algorithm that combined NLP, vital signs, and laboratory values had a higher sensitivity than NLP alone, which was used in another study (95% v 88%).³⁰ In common among these studies is the improved disease prediction by combining multiple sources of data (structured and unstructured) with NLP, and, indeed, our study supports this approach.

Our study has several limitations. The modest data set of 64 case patients was a major limitation, but emblematic of the difficulties acquiring large training data sets for a disease for which routine epidemiologic surveillance does not exist. The probabilities from the text classifier were derived from our earlier work¹¹ rather than acquired in a pipeline approach with sequential classifiers operating; however, this should not have appreciably changed our overall findings. Our case patients did not exclusively have proven or probable IMD (despite the fact that these represented a higher degree of certainty) for several reasons: possible IMD cases (those with radiologic features but lacking positive microbiology) represent a substantial burden in clinical practice and may predominate (up to 90%) in some centers^{2,31}; possible IMD cases are treated similarly to proven and/or probable IMD cases and consume equivalent health care resources (eg, diagnostics, antifungal drug use),¹ and their exclusion would underestimate the true prevalence of IMD. Notably, all case patients underwent expert adjudication in accordance with international definitions.¹⁴ We focused on the highest-risk population, and limited generalizability to other risk groups (eg, solid organ transplant). Finally, our data set was enriched with positive cases, but external prospective validation of the expert system in clinical practice where prevalence rates are lower, and among diverse patient groups, will be the focus of future work.

A disease with an evolving epidemiology associated with the introduction of novel cancer therapeutics, expanded transplantation services, an aging population, and neglected groups such as children, requires a surveillance system with the agility to keep pace with change. An electronic surveillance system could meet this demand, particularly with the increasing digitization of health care. Our expert system using the wealth of electronic data available in hospitals could be used to flag patients with IMD for further review. The addition of other data analytics to our

Table 5. Error Log of False Negatives and False Positives for Invasive Mold Diseases From the Expert System Incorporating the Naive Bayes Algorithm

| Patient | Output of Text Classifier | | | Microbiology Investigation | | | | | | Antifungal Drug Rule ¹ | | | Prediction by Expert System |
|---|---------------------------|--------------------|------------------|----------------------------|------------------|-----------------|----------------------|--------------------------|----------|-----------------------------------|--------|-----|-----------------------------|
| | Scan_Any positive | Blood_Any positive | BAL_Any positive | Fluid_Any positive | PCR_Any positive | GM_Any positive | Sterile_Any positive | Non_Sterile_Any positive | Drug 1 | Drug 2 | Drug 3 | | |
| False negatives (ie, fungal cases classified as negative) | | | | | | | | | | | | | 0 |
| 1 | 0 | No test | No test | No test | No test | No test | No test | No test | No test | low | low | No | 0 |
| False positives (ie, negative cases classified as fungal cases) | | | | | | | | | | | | | |
| 1 | 0.71 | No test | No test | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.56 |
| 2 | 0.83 | Negative | Negative | Negative | No test | No test | No test | Negative | Negative | No | No | No | 0.81 |
| 3 | 0.86 | No test | No test | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.74 |
| 4 | 0.71 | Negative | No test | No test | Negative | Negative | No test | positive | positive | Yes | No | No | 1.00 |
| 5 | 1.00 | Negative | No test | No test | No test | No test | No test | No test | No test | Low | No | No | 0.71 |
| 6 | 0.75 | Negative | No test | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.66 |
| 7 | 0.67 | Negative | Negative | No test | No test | No test | No test | No test | No test | Low | Low | No | 0.77 |
| 8 | 1.00 | No test | No test | No test | No test | No test | No test | No test | No test | No | No | No | 0.39 |
| 9 | 1.00 | Negative | Negative | No test | Negative | No test | No test | No test | No test | Yes | Low | No | 0.98 |
| 10 | 0.75 | Negative | Negative | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.83 |
| 11 | 1.00 | Negative | No test | No test | No test | No test | No test | No test | No test | Low | No | No | 0.73 |
| 12 | 0.71 | Negative | No test | No test | No test | No test | No test | No test | No test | Low | No | No | 0.43 |
| 13 | 0.80 | No test | No test | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.69 |
| 14 | 0.67 | No test | No test | No test | No test | No test | No test | No test | No test | Yes | Low | Low | 0.82 |
| 15 | 0.67 | Negative | No test | No test | No test | No test | No test | No test | No test | Yes | No | No | 0.41 |

Abbreviations: BAL, bronchoalveolar lavage; GM, galactomannan; PCR, polymerase chain reaction.

expert system, such as image analysis of CT scans, is an area of active research³² that we hope will improve the performance of the expert system and provide decision support for a range of activities, including radiologic

diagnosis and antifungal stewardship programs in hospitals.⁸

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